

· 综述 ·

CTRP 家族与糖尿病及其并发症

徐佳佳 张慧娟

哈尔滨医科大学附属第一医院内分泌科 150000

通信作者:张慧娟,Email:hydzjh@126.com

【摘要】 补体 C1q/肿瘤坏死因子相关蛋白(CTRP)家族是近来新发现的脂肪因子,与脂联素具有高度同源性。CTRP 家族在糖、脂代谢、能量代谢、炎症反应调控、血管保护中发挥重要作用。近年来研究发现,CTRP 家族可以调控糖代谢、影响胰岛素抵抗等过程,参与糖尿病及其并发症的发生、发展,并有望为糖尿病及其并发症提供新的靶点。

【关键词】 CTRP 蛋白;糖尿病;糖尿病并发症

DOI:10.3760/cma.j.issn.1673-4157.2019.02.010

CTRP family in diabetes mellitus and its complications Xu Jiajia, Zhang Huijuan. Department of Endocrinology, The First Affiliated Hospital of Harbin Medical University, Harbin 150000, China

Corresponding author: Zhang Huijuan, Email:hydzjh@126.com

【Abstract】 Complement-C1q/tumor necrosis factor-related protein (CTRP) family is a recently discovered adipokine, which is highly homologous with adiponectin. CTRP family plays an important role in the regulation of glycolipid metabolism, energy metabolism, inflammatory response and vascular protection. In recent years, the researchers found that CTRP family members can participate in the development of diabetes and its complications by regulating glucose metabolism, affecting insulin resistance, and is expected to provide new therapeutic targets for diabetes and its complications.

【Key words】 Complement-C1q/tumor necrosis factor-related protein; Diabetes mellitus; Diabetic complications

DOI:10.3760/cma.j.issn.1673-4157.2019.02.010

脂肪组织不仅是机体能量的储存器官,而且是一个重要的内分泌器官,分泌和表达多种蛋白质,如脂联素、抵抗素、瘦素等^[1]。新近研究表明,补体 C1q/肿瘤坏死因子相关蛋白(CTRP)属于脂肪细胞因子超家族,可以直接或间接的调节葡萄糖代谢,影响胰岛素敏感性,调控炎症反应,与糖尿病及其并发症密切相关。

1 CTRP 家族成员的结构与组织表达

CTRP 家族由一组高度保守,与脂联素结构相似的蛋白质组成^[2]。其属于 C1q 家族的小分泌蛋白,共包括 15 个成员,分别为 CTRP 1~15,每个成员都有一个共同的结构,分别由氨基末端的信号肽、一个短的可变区域、一个胶原样结构域和一个羧基末端的球形结构域组成,其中羧基末端球形结构域是发挥生物学功能的重要结构。各个成员均可形成同源三聚体,多数成员可形成同源六聚体或高分子量的多聚体。CTRP 蛋白家族在组织表达中存在差

异,其中,脂肪组织中表达 CTRP 家族种类最多,包括 CTRP1~7、CTRP9、CTRP11~13。肺和睾丸主要表达 CTRP7、CTRP8,胎盘主要表达 CTRP1、CTRP3、CTRP6、CTRP10,心脏主要表达 CTRP1、CTRP9,软骨细胞主要表达 CTRP3,同时发现 CTRP5 在人视网膜色素上皮细胞特异性表达。

2 CTRP 家族与糖尿病

2.1 CTRP 家族与糖代谢 CTRP 家族成员通过调控糖代谢,降低血糖。研究显示,重组 CTRP1 可激活分化的小鼠肌管中蛋白激酶 B(Akt)和 p44/42 丝裂原活化蛋白激酶(MAPK)信号通路,Akt 的激活增加了葡萄糖转运蛋白 4 向质膜的转运,从而增强肌肉组织对葡萄糖的吸收^[3]。Peterson 等^[4]在培养大鼠 H4IIE 肝细胞中发现,重组 CTRP1 并没有改变 AMPK 信号途径、葡萄糖输出、糖异生基因表达,而是激活骨骼肌中 AMPK 信号途径,使乙酰辅酶 A 羧化酶磷酸化,增强脂肪酸氧化和能量消耗。以上研究

表明,肌肉组织可能是 CTRP 家族成员降糖的作用靶点,并且直接影响肌肉组织中的信号通路和糖代谢。

临床试验发现,与对照组相比,糖尿病前期者与 2 型糖尿病患者血浆 CTRP3 水平明显降低^[5],表明 CTRP3 可能与血糖有密切联系。相关动物实验也证实了这一点, Peterson 等^[6]通过重组蛋白 CTRP3 给药,发现野生型小鼠和肥胖 ob/ob 小鼠中血浆 CTRP3 水平增加 3 倍,血糖水平明显降低,并且未影响胰岛素、胰高血糖素和脂联素水平;其原因是 CTRP3 激活肝脏中 Akt 信号通路,抑制肝脏糖异生酶的表达。表明 CTRP3 作为一种新的脂肪因子,在糖代谢中起重要作用。另外研究发现,过表达 CTRP9 可以降低 ob/ob 小鼠血糖水平,同时,CTR9 可以刺激分化的肌管中 AMPK、Akt 和 p44/42 MAPK 磷酸化^[7]。CTR12 可通过激活磷脂酰肌醇 3 激酶 (PI3K)-Akt 信号通路,抑制肝细胞糖异生,促进脂肪细胞对葡萄糖的摄取,增强糖尿病小鼠脂肪组织和肝脏的胰岛素信号转导。以上均表明 CTRP 家族成员作为新的脂肪因子,可以通过多种机制调节糖代谢,将可能成为新的一种抗糖尿病因子^[8]。

但 Bai 等^[9]最新临床相关试验显示,与对照组相比,2 型糖尿病患者血清 CTRP1 水平明显升高,差异具有统计学意义,而 CTRP9 与 CTRP13 无明显差异;行口服葡萄糖耐量试验 2 h 后,血清 CTRP1 水平显著升高,这些脂肪因子在糖尿病患者中没有降低反而升高,可能为机体保护机制的反馈调节使患者脂肪组织合成增多,亦有可能存在其他因素影响 CTRP 家族水平,故 CTRP 与糖尿病的机制需要进一步研究。

2.2 CTRP 家族与胰岛素抵抗 Xin 等^[10]发现,CTR1 不仅增加脂肪细胞对葡萄糖的利用率,并且通过作用于胰岛素受体底物 (IRS)-1 Ser¹¹⁰¹ 的磷酸化,降低胰岛素抵抗。动物实验表明,CTR1 过表达可以增加外周组织胰岛素敏感性,提示 CTR1 在胰岛素抵抗中发挥重要作用^[11]。相关临床试验发现,糖尿病患者循环 CTRP1 水平与胰岛素抵抗程度呈负相关^[12]。

CTR3 可以通过减轻炎症反应和改善胰岛素信号转导而改善胰岛素抵抗。研究人员将重组 CTR3 作用于胰岛素抵抗的 3T3-L1 脂肪细胞,结果白细胞介素-6 和肿瘤坏死因子- α 的释放减少,抑制炎症反应,并且增加胰岛素敏感性;此外,CTR3 还

增加了 PI3K 和 Akt 及葡萄糖转运蛋白 4 mRNA 的表达,从而改善胰岛素信号转导,提高胰岛素敏感性^[13]。这些研究表明 CTR3 直接影响脂肪细胞内信号通路,抑制脂肪组织炎症反应,对抗胰岛素抵抗的发生。

另有实验表明,尽管在肥胖小鼠模型中 CTR12 mRNA 水平受到抑制,但给予抗糖尿病药物罗格列酮后可上调该因子的表达,提示其在促进胰岛素敏感性方面可能起作用。给予腺病毒载体介导的 CTR12 可降低肥胖小鼠脂肪组织中巨噬细胞的积累和促炎基因的表达^[14]。表明 CTR12 可以通过抑制脂肪组织炎症反应,阻止胰岛素抵抗的发生,增加胰岛素敏感性,对糖尿病的防治可能起到关键性作用。

3 CTRP 家族与糖尿病并发症

3.1 CTRP 家族与糖尿病大血管并发症 以往的研究提示,CTR9 对大血管具有保护作用^[15]。已证实 CTR9 可减轻糖尿病小鼠主动脉内皮细胞活性氧簇的产生,抑制氧化应激,且增加内源性抗氧化酶的表达,提示 CTR9 对高糖诱导的氧化应激具有保护作用^[16]。线粒体生物合成已被发现有助于减轻氧化应激作用^[17]。研究报道,在高糖处理的内皮细胞线粒体生物合成过程中,过氧化物酶体增殖物活化受体 γ 协同刺激因子-1 α (PGC-1 α) 是一种重要的辅激活因子,PGC-1 α 在内皮细胞中的过度表达可上调线粒体膜电位和抑制细胞凋亡^[18-19]。沉默信息调节因子 1 (SIRT1) 介导的 PGC-1 α 去乙酰化是激活线粒体脂肪酸氧化基因所必需的^[20]。Cheng 等^[16]表明,CTR9 可以通过上调 PGC-1 α 和 SIRT1 表达,增加线粒体的合成,提示 CTR9 可以减轻氧化应激,增加线粒体合成从而保护内皮细胞功能。

3.2 CTRP 家族与糖尿病微血管并发症 临床研究显示,在糖尿病视网膜病变非增殖期患者玻璃体中血管内皮细胞黏附因子-1 (VCAM-1) 水平升高,而 VCAM-1 水平可能反映内皮细胞损伤^[21-22]。Yan 等^[23]发现,CTR3 可以通过激活 AMPK 信号通路,抑制在高糖诱导下视网膜微血管内皮细胞 VCAM-1 的表达,ELISA 法检测人血清中 CTRP3 含量,与对照组相比,糖尿病视网膜病变非增殖期与增殖期患者血清 CTRP3 水平逐渐降低,提示 CTR3 对糖尿病视网膜病变的保护作用,其机制可能为 CTR3 通过激活 AMPK 通路抑制 VCAM 堆积,保护视网膜微血管内皮细胞。糖尿病心肌病是糖尿病患者心力衰竭的主要诱因^[24]。研究发现,在糖尿病大鼠心肌细胞中

CTRP3 表达明显降低,敲除CTRP3 基因后发现,心肌细胞氧化应激与炎症因子表达增加,心肌细胞存活率降低。通过链脲佐菌素诱导糖尿病大鼠心脏损伤和心功能减退模型,发现CTRP3 可以改善糖尿病引起的心脏损伤与心功能不全^[25]。结果表明,CTRP3 具有治疗糖尿病心肌病变的潜力,机制为通过激活 AMPK 通路,抑制氧化应激与炎症因子的表达,减少心肌细胞死亡,保护心功能。这也为治疗糖尿病心肌病提供了一种很有前景的方法。

综上所述,CTRP 家族成员作为新发现的脂肪因子,与糖尿病以及糖尿病并发症存在密切关系。但现在的研究仍然十分局限,其生物作用需要更多临床试验支持。因此,深入了解 CTRP 家族将有助于加深对疾病的了解。

参 考 文 献

- [1] Trujillo ME, Scherer PE. Adipose tissue-derived factors: impact on health and disease[J]. *Endocr Rev*, 2006, 27(7): 762-778. DOI: 10.1210/er.2006-0033.
- [2] Schäffler A, Buechler C. CTRP family: linking immunity to metabolism[J]. *Trends Endocrinol Metab*, 2012, 23(4): 194-204. DOI: 10.1016/j.tem.2011.12.003.
- [3] Wong GW, Krawczyk SA, Kitidis-Mitrokostas C, et al. Molecular, biochemical and functional characterizations of C1q/TNF family members: adipose-tissue-selective expression patterns, regulation by PPAR- γ agonist, cysteine-mediated oligomerizations, combinatorial associations and metabolic functions[J]. *Biochem J*, 2008, 416(2): 161-177. DOI: 10.1042/BJ20081240.
- [4] Peterson JM, Aja S, Wei Z, et al. CTRP1 protein enhances fatty acid oxidation via AMP-activated protein kinase (AMPK) activation and acetyl-CoA carboxylase (ACC) inhibition[J]. *J Biol Chem*, 2012, 287(2): 1576-1587. DOI: 10.1074/jbc.M111.278333.
- [5] Wei H, Qu H, Wang H, et al. Plasma C1q/TNF-Related Protein-3 (CTRP-3) and High-Mobility Group Box-1 (HMGB-1) concentrations in subjects with prediabetes and type 2 diabetes[J]. *J Diabetes Res*, 2016, 2016: 9438760. DOI: 10.1155/2016/9438760.
- [6] Peterson JM, Wei Z, Wong GW. C1q/TNF-related protein-3 (CTRP3), a novel adipokine that regulates hepatic glucose output[J]. *J Biol Chem*, 2010, 285(51): 39691-39701. DOI: 10.1074/jbc.M110.180695.
- [7] Wong GW, Krawczyk SA, Kitidis-Mitrokostas C, et al. Identification and characterization of CTRP9, a novel secreted glycoprotein, from adipose tissue that reduces serum glucose in mice and forms heterotrimer with adiponectin[J]. *FASEB J*, 2009, 23(1): 241-258. DOI: 10.1096/fj.08-114991.
- [8] Wei Z, Peterson JM, Lei X, et al. C1q/TNF-related protein-12 (CTRP12), a novel adipokine that improves insulin sensitivity and glycemic control in mouse models of obesity and diabetes[J]. *J Biol Chem*, 2012, 287(13): 10301-10315. DOI: 10.1074/jbc.M111.303651.
- [9] Bai B, Ban B, Liu Z, et al. Circulating C1q complement/TNF-related protein (CTRP) 1, CTRP9, CTRP12 and CTRP13 concentrations in type 2 diabetes mellitus: *in vivo* regulation by glucose[J]. *PLoS One*, 2017, 12(2): e0172271. DOI: 10.1371/journal.pone.0172271.
- [10] Xin Y, Zhang D, Fu Y, et al. C1q/TNF-related protein 1 improve insulin resistance by reducing phosphorylation of serine 1101 in insulin receptor substrate 1[J]. *Endocr J*, 2017, 64(8): 787-796. DOI: 10.1507/endocrj.EJ17-0128.
- [11] Peterson JM, Aja S, Wei Z, et al. CTRP1 protein enhances fatty acid oxidation via AMP-activated protein kinase (AMPK) activation and acetyl-CoA carboxylase (ACC) inhibition[J]. *J Biol Chem*, 2012, 287(2): 1576-1587. DOI: 10.1074/jbc.M111.278333.
- [12] Pan X, Lu T, Wu F, et al. Circulating complement-C1q TNF-related protein 1 levels are increased in patients with type 2 diabetes and are associated with insulin sensitivity in Chinese subjects[J]. *PLoS One*, 2014, 9(5): e94478. DOI: 10.1371/journal.pone.0094478.
- [13] Li X, Jiang L, Yang M, et al. CTRP3 improves the insulin sensitivity of 3T3-L1 adipocytes by inhibiting inflammation and ameliorating insulin signalling transduction[J]. *Endokrynol Pol*, 2014, 65(4): 252-258. DOI: 10.5603/EP.2014.0034.
- [14] Enomoto T, Ohashi K, Shibata R, et al. Adiponin/C1qdc2/CTRP12 protein functions as an adipokine that improves glucose metabolism[J]. *J Biol Chem*, 2011, 286(40): 34552-34558. DOI: 10.1074/jbc.M111.277319.
- [15] Sun Y, Yi W, Yuan Y, et al. C1q/tumor necrosis factor-related protein-9, a novel adipocyte-derived cytokine, attenuates adverse remodeling in the ischemic mouse heart via protein kinase A activation[J]. *Circulation*, 2013, 128(11 Suppl 1): S113-S120. DOI: 10.1161/CIRCULATIONAHA.112.000010.
- [16] Cheng L, Li B, Chen X, et al. CTRP9 induces mitochondrial biogenesis and protects high glucose-induced endothelial oxidative damage via AdipoR1-SIRT1-PGC-1 α activation[J]. *Biochem Biophys Res Commun*, 2016, 477(4): 685-691. DOI: 10.1016/j.bbrc.2016.06.120.
- [17] de Oliveira MR, Nabavi SF, Manayi A, et al. Resveratrol and the mitochondria: from triggering the intrinsic apoptotic pathway to inducing mitochondrial biogenesis, a mechanistic view[J]. *Biochim Biophys Acta*, 2016, 1860(4): 727-745. DOI: 10.1016/j.bbagen.2016.01.017.
- [18] Jiang Y, Xia W, Yang J, et al. BPA-induced DNA hypermethylation of the master mitochondrial gene PGC-1 α contributes to cardiomyopathy in male rats[J]. *Toxicology*, 2015, 329: 21-31. DOI: 10.1016/j.tox.2015.01.001.
- [19] Scarpulla RC. Metabolic control of mitochondrial biogenesis through the PGC-1 family regulatory network[J]. *Biochim Biophys Acta*, 2011, 1813(7): 1269-1278. DOI: 10.1016/j.bbamer.2010.09.

- 019.
- [20] Yu L, Liang H, Dong X, et al. Reduced silent information regulator 1 signaling exacerbates myocardial ischemia-reperfusion injury in type 2 diabetic rats and the protective effect of melatonin[J]. J Pineal Res, 2015, 59(3): 376-390. DOI:10.1111/jpi.12269.
- [21] Limb GA, Hickman-Casey J, Hollifield RD, et al. Vascular adhesion molecules in vitreous from eyes with proliferative diabetic retinopathy[J]. Invest Ophthalmol Vis Sci, 1999, 40(10): 2453-2457.
- [22] Yoshizawa M, Nagai Y, Ohsawa K, et al. Elevated serum levels of soluble vascular cell adhesion molecule-1 in NIDDM patients with proliferative diabetic retinopathy[J]. Diabetes Res Clin Pract, 1998, 42(1): 65-70.
- [23] Yan Z, Zhao J, Gan L, et al. CTRP3 is a novel biomarker for diabetic retinopathy and inhibits HGHL-induced VCAM-1 expression in an AMPK-dependent manner [J]. PLoS One, 2017, 12(6): e0178253. DOI:10.1371/journal.pone.0178253.
- [24] Bugger H, Abel ED. Molecular mechanisms of diabetic cardiomyopathy[J]. Diabetologia, 2014, 57(4): 660-671. DOI:10.1007/s00125-014-3171-6.
- [25] Ma ZG, Yuan YP, Xu SC, et al. CTRP3 attenuates cardiac dysfunction, inflammation, oxidative stress and cell death in diabetic cardiomyopathy in rats[J]. Diabetologia, 2017, 60(6): 1126-1137. DOI: 10.1007/s00125-017-4232-4.

(收稿日期:2018-07-25)

(本文编辑:饶颖)

(上接第 111 页)

- [11] Hansen TB, Jensen TI, Clausen BH, et al. Natural RNA circles function as efficient microRNA sponges [J]. Nature, 2013, 495(7441): 384-388. DOI:10.1038/nature11993.
- [12] Li Z, Huang C, Bao C, et al. Exon-intron circular RNAs regulate transcription in the nucleus [J]. Nat Struct Mol Biol, 2015, 22(3): 256-264. DOI:10.1038/nsmb.2959.
- [13] Li X, Liu CX, Xue W, et al. Coordinated circRNA biogenesis and function with NF90/NF110 in viral infection[J]. Mol Cell, 2017, 67(2): 214-227. e7. DOI:10.1016/j.molcel.2017.05.023.
- [14] Du WW, Yang W, Chen Y, et al. Foxo3 circular RNA promotes cardiac senescence by modulating multiple factors associated with stress and senescence responses[J]. Eur Heart J, 2017, 38(18): 1402-1412. DOI:10.1093/eurheartj/ehw001.
- [15] Zhao Z, Li X, Jian D, et al. Hsa_circ_0054633 in peripheral blood can be used as a diagnostic biomarker of pre-diabetes and type 2 diabetes mellitus [J]. Acta Diabetol, 2017, 54(3): 237-245. DOI:10.1007/s00592-016-0943-0.
- [16] Shang FF, Luo S, Liang X, et al. Alterations of circular RNAs in hyperglycemic human endothelial cells[J]. Biochem Biophys Res Commun, 2018, 499(3): 551-555. DOI:10.1016/j.bbrc.2018.03.187.
- [17] Yan L, Feng J, Cheng F, et al. Circular RNA expression profiles in placental villi from women with gestational diabetes mellitus[J]. Biochem Biophys Res Commun, 2018, 498(4): 743-750. DOI: 10.1016/j.bbrc.2018.03.051.
- [18] Stoll L, Sobel J, Rodriguez-Trejo A, et al. Circular RNAs as novel regulators of β -cell functions in normal and disease conditions [J]. Mol Metab, 2018, 9: 69-83. DOI: 10.1016/j.molmet.2018.01.010.
- [19] Xu H, Guo S, Li W, et al. The circular RNA Cdr1as, via miR-7 and its targets, regulates insulin transcription and secretion in islet cells[J]. Sci Rep, 2015, 5: 12453. DOI:10.1038/srep12453.
- [20] Fang Y, Wang X, Li W, et al. Screening of circular RNAs and validation of circANKRD36 associated with inflammation in patients with type 2 diabetes mellitus[J]. Int J Mol Med, 2018, 42(4): 1865-1874. DOI:10.3892/ijmm.2018.3783.
- [21] Gu Y, Ke G, Wang L, et al. Altered expression profile of circular RNAs in the serum of patients with diabetic retinopathy revealed by microarray[J]. Ophthalmic Res, 2017, 58(3): 176-184. DOI: 10.1159/000479156.
- [22] Zhang SJ, Chen X, Li CP, et al. Identification and characterization of circular RNAs as a new class of putative biomarkers in diabetes retinopathy[J]. Invest Ophthalmol Vis Sci, 2017, 58(14): 6500-6509. DOI:10.1167/iovs.17-22698.
- [23] Shan K, Liu C, Liu BH, et al. Circular noncoding RNA HIPK3 mediates retinal vascular dysfunction in diabetes mellitus[J]. Circulation, 2017, 136(17): 1629-1642. DOI:10.1161/CIRCULATIONAHA.117.029004.
- [24] Hu W, Han Q, Zhao L, et al. Circular RNA circRNA_15698 aggravates the extracellular matrix of diabetic nephropathymesangial cells via miR-185/TGF- β 1 [J]. J Cell Physiol, 2019, 234(2): 1469-1476. DOI:10.1002/jcp.26959.
- [25] Zhou B, Yu JW. A novel identified circular RNA, circRNA_010567, promotes myocardial fibrosis via suppressing miR-141 by targeting TGF- β 1 [J]. Biochem Biophys Res Commun, 2017, 487(4): 769-775. DOI:10.1016/j.bbrc.2017.04.044.
- [26] Wang L, Luo T, Bao Z, et al. Intrathecal circHIPK3 shRNA alleviates neuropathic pain in diabetic rats [J]. Biochem Biophys Res Commun, 2018, 505(3): 644-650. DOI:10.1016/j.bbrc.2018.09.158.

(收稿日期:2018-08-16)

(本文编辑:饶颖)